# **RESEARCH ARTICLE**

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# Degradation and Microbiological Validation of Meropenem Antibiotic in Aqueous Solution Using UV, $UV/H_2O_2$ , $UV/TiO_2$ and $UV/TiO_2/H_2O_2$ Processes.

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# ABSTRACT

Aqueous UV, UV/H<sub>2</sub>O<sub>2</sub>, UV/TiO<sub>2</sub> and UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub>mediateddegradation/oxidation of the carbapenem antibiotic, meropenem (MERO) was experimentally studied. Degussa P-25 titanium dioxide was used as photocatalyst and UV-light source was used for activation of TiO<sub>2</sub>.The nanosized titanium dioxide was immobilized on the glass support for improving the efficiency and economics of the photocatalytic processes. The immobilized film of titanium dioxide has been characterized, using X-ray diffraction (XRD) andscanning electron microscopy (SEM).The study of antibiotic degradation was conducted in the specific Batch Photocatalytic Reactor. MERO standard solution was used at 500  $\mu$ g/ml concentration, which degraded up to 99% of antibiotics. Microbiological assay showed that the loss of antibacterial activity is directly proportional to the time of UV-irradiation. The experiment also showed that the UV-irradiation itself causes the degradation of antibiotics, but in very slow manner in comparison to the photocatalysis process. The experimental study showed that UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system is effective and efficient for the treatment of antibiotic waste. *Keywords*- Photocatalysis, Meropenem, Antibiotics, TiO<sub>2</sub>, UV-irradiation.

#### I. INTRODUCTION

Carbapenem are a class of β-lactam broadspectrum antibacterial agents used for human and veterinary application. It act by inhibiting the cell wall synthesis and are known to be most effective against gram negative infections. As a result of their rising popularity, overuse and misuse of antibiotics has fuelled a rise in drug-resistant infections and experts are particularly alarmed about bacteria that cannot be killed with carbapenems[1-3], the foremost powerful category of antibiotics. Being persistent compounds[4-6], antibiotics pass the several treatment plants intact and accumulate in the environment[7-11]. Higher concentration of these compounds in the environment causes damage to the micro-flora and fauna [12,13], and its lower concentration in the environment promotes the development of antibiotic resistant bacteria[14-17].

In recent decades, the emergence of antibiotic resistant bacteria has increased and many specialists believe that the increase is due to the inappropriate use of antibiotics [18, 19]. Furthermore, the presence of antibiotics in aqueous effluent has also increased and their removal will be a challenge in the near future.

Therefore, physical-chemical removal technologies are needed such as advanced oxidation processes (AOPs) like photolysis, photocatalysis andincorporation of strong oxidizing agents like H<sub>2</sub>O<sub>2</sub>. Among the different AOPs, TiO<sub>2</sub>photocatalysis emerges as a promising treatment technology due to its specific advantages like lack of mass transfer limitations, operation at ambient conditions and catalyst is inexpensive, commercially accessible, non-toxic and photochemically stable. One of the most widely used carbapenem antibiotics is meropenem (MERO,(4R,5S,6S)-3-[(3S,5S) -5-(dimethylcarbamoyl)pyrrolidin -3-yl]sulfanyl -6-[(1R) - 1 - hydroxyethyl] - 4 - methyl - 7 - oxo - 1- azabicyclo[3.2.0]hept - 2 - ene-2-carboxylic acid, see Fig. 1). To the best of author's knowledge, there are no works dealing with the AOPsapplication to the degradation. application MERO The of microbiological assay for the determination of MERO concentration in its treated solutions makes the novelty of the present research.

The UV/H<sub>2</sub>O<sub>2</sub>, UV/TiO<sub>2</sub> and UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub>processes are the UV based advanced oxidation processes (AOPs) and generates the hydroxyl radical ( $^{\circ}$ OH) which has strong oxidizing capabilities [20]. Many studies have illustrated the effectiveness of these processes in the oxidation and mineralization of various pharmaceutical compounds and these processes has been widely studied for the remediation of both ground and drinking waters [21-25].



Fig. 1.Meropenem (MERO) molecule.

Although these AOPs are efficient in the mineralization of various antibiotics[26-50], however, it is uncertain whether carbapenemantibiotics like MERO can be completely mineralized by these processes. It is also unknown whether the process renders MERO and its potential photocatalytictransformation products biologically inactive, as bioactivity is an important parameter to consider when assessing the feasibility of an advanced degradation system.

The aim of this study is to investigate the elimination of MERO and look at the changes in its antibacterial activity using UV,  $UV/H_2O_2$ ,  $UV/TiO_2$  and  $UV/TiO_2/H_2O_2$  processes.Microbial growth inhibition bioassay will be performed to determine the loss of antibacterial activity as well as concentration of intact MERO in treated solutions. In this study, we also examine and compare the degradation kinetics for these processes.

# **II. MATERIALS AND METHODS**

#### 2.1 Chemicals

Meropenem, ADH2031, was provided by Cipla (India). Hydrogen peroxide (> 30%) was obtained Merck (India). Ethanol (99.9%) from and Whatmanfilter paper no.-1 were purchased from s d Fine Chem Ltd (India). Mueller Hinton agar and LB broth media, used for microbiological assay, were obtained from HIMEDIA (India).Degussa P25 TiO<sub>2</sub>; commercial photocatalyst provided by Evonik Industries, Germany was used throughout the current investigation. According to manufacturer's specifications, its BET surface area was  $50\pm10 \text{ m}^2/\text{g}$ , average particle size was around 25 nm, purity was 99.5 %, and anatase to rutile ratio was 3:1. Any other chemicals used in this study were of analytical grade. Stock solution of MERO and other solution were prepared in distilled water (DW) and diluted as required.

# 2.2 Coating of Degussa P25 film on petri-plate by Dip-Coating method

The catalyst (TiO<sub>2</sub>) was immobilized on the petri plate using Dip-Coating method according to the process mentioned by Negishi et al. 1998 [51], with some modifications. The TiO<sub>2</sub> suspension was prepared by adding 2gm of TiO<sub>2</sub> in 40 ml of distilled water in a beaker. The resulting solution was stirred continuously for 15-20 min until all the TiO<sub>2</sub> powder gets mixed up properly. After the suspension was prepared, the petri-plates (100 mm diameter) were dipped in a beaker containing the suspension for 2-3 min and again taken out from the suspension (i.e. simply consider it as one time dip-coating). After dipcoating, the plates were dried in an oven for 1 hour at 108°C. If layer deposited on the plates was not uniform, then plates were dipped again in suspension to obtain uniform layer and above procedure was repeated. After drying, the plates were carried to muffle furnace where the coated supports were annealed for 3-4 hours at 500°C. Annealing provides better mechanical strength to films. Coated petriplates having active catalyst surface area of 78.54 cm<sup>2</sup> were used as such for photocatalytic experiments.

#### 2.3 Characterization of coated films

The morphology of the coated titanium dioxide was studied using SEM. The test sample was analyzed in the scanning electron microscope JSM 6100 (JEOL), which operated at 25 kV.The XRD analysis of the titanium dioxide coating was done by the plate XRD technique to examine the crystalline structure of it. The X-ray diffraction pattern was established on a Phillips PW-1710 X-ray diffractometer using Cu-K $\alpha$  radiation as X-ray source at an angle of  $2\theta$  ranging from 20° to 80°. The scanning rate of 0.034( $2\theta$ )/s was used for the measurements.

# **2.4 Experimental procedure**

Photocatalytic degradation of MERO was carried out in a Batch Photocatalytic Reactor (BPR) at room temperature ( $23 \pm 2$  °C), manufactured by LabcoIndia limited, having dimension (52cm x 28cm x 56cm). The reaction chamber contains two UV Tube light (Philips, TUV, 15W/G15 T8) placed at the top of the reactor, one exhaust fan (to control inside temperature) fitted at the back side of the reactor, a temperature probe fitted at the top of reactor, a magnetic stirrer placed at the bottom of reactor with a height adjustable stand as shown in Fig. 2. Inner walls of the reactor were fully coated with silver paint in order to minimize the irradiation losses. A MERO solution of 20 ml with concentration of 500 µg/ml was poured into petri plates having a similar cross-sectional area of  $78.54 \text{ cm}^2$ .



Four different modes of treatment were studied for the degradation of MERO solution. UV irradiation and operating conditions were similar for each mode of degradation. But, they differ only in the presence of the TiO<sub>2</sub> as photocatalyst and - H<sub>2</sub>O<sub>2</sub> as strong oxidizing agent. UV/TiO<sub>2</sub> system involves UV-irradiation in the presence of TiO<sub>2</sub> photocatalyst, MERO solution was poured into a TiO<sub>2</sub> coated petri plate; UV system uses UV-irradiation only, MERO solution was poured into an uncoated petri plate: UV/H<sub>2</sub>O<sub>2</sub> system uses UV-irradiation in the presence of hydrogen peroxide, MERO solution was poured into an uncoated petri plate containing H<sub>2</sub>O<sub>2</sub> (100 mg/L); UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system involves the combination of UV-irradiation, TiO<sub>2</sub> photocatalyst and oxidizing agent hydrogen peroxide. MERO solution was poured into a TiO<sub>2</sub> coated petri plate containing H<sub>2</sub>O<sub>2</sub> (100 mg/L). These plates were placed into the reaction chamber of BPR for degradation. During the experiment, aliquots of 0.2 ml of MERO solution were withdrawn from the petri plates at regular intervals to observe the degradation of MERO. The aliquots withdrawn were then analyzed by Microbiological assay.

#### 2.5 Microbiological assay

The antibacterial activity of the MERO solution, before, during and after treatment by UV, UV/H<sub>2</sub>O<sub>2</sub>, UV/TiO<sub>2</sub> and UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> processes were measured using non-pathogenic bacterial strain, *E. coli* MTCC 443 as a test microorganism, according to the protocol based on Disk diffusion method [52]. In brief, 0.2 ml of standard solution of MERO and the solution treated with UV, UV/H<sub>2</sub>O<sub>2</sub>, UV/TiO<sub>2</sub> and

UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> processes were taken. From the collected samples, 7µL aliquots were used for the impregnation of filter paper discs having diameter of 5 mm. The test microorganism was cultured in LB broth, adjusting its concentration to the 0.5 McFarland standards, containing approximately  $1.5 \times 10^8$  CFU/mL. The dried surface of a Mueller Hinton agar plates were inoculated by streaking the swab over the entire sterile agar surface. The antibiotic impregnated discs were dispensed onto the surface of the inoculated agar plate. In addition, the positive and negative growth controls were also used. The plates were incubated at 37 °C for 16 hours. Each assay was realized in triplicate. After the incubation, the inhibition zones (halos) were measured and correlated with MERO degradation.

# III. RESULTS AND DISCUSSION

**3.1** Characterization of  $TiO_2$  coated film

The XRD studies were made to notice any change in the phase composition and photocatalytic properties of the titanium dioxide after immobilization on the plate. The XRD measurements were done directly on coated catalyst resulting in noisy patterns. The XRD pattern of TiO<sub>2</sub> coated film is presented in Fig. 3. The XRD patterns of the coated TiO<sub>2</sub> surface did not show any variation in the structure and phase composition due to coating process. The diffraction peaks observed at  $2\theta$  = 25.31°, 37.91°, 48.15°, 53.92°, 55.16°, 62.84°, 69.12°, 70.41° and 75.02° correspond to the known diffraction maxima of anatase phase of TiO<sub>2</sub>. The peaks at  $2\theta = 27.45^{\circ}$ ,  $36.10^{\circ}$ ,  $41.66^{\circ}$ ,  $54.34^{\circ}$ , and 56.61° correspond to rutile phase of TiO<sub>2</sub>. The intensity of the peaks reveals that the major phase available in the coated TiO<sub>2</sub> is anatase while the rutile is the minor phase. The anatase TiO<sub>2</sub> is necessary to achieve the required electronic band gap of 3.2 eV.The photocatalytic reaction is very efficient on the mixture of anatase and rutile particles [53].

To study the morphology of  $\text{TiO}_2$  coating, the SEM images of the prepared film were taken at  $\times 10,000$  magnifications, and shown in Fig. 4. SEM image demonstrate the nanostructure of the catalyst, and rough surface of the film, which is quite necessary for the good photocatalytic activity of the film. It is obvious from the micrographs that TiO<sub>2</sub> (white spots) is more uniformly distributed on the film surface, which indicate the availability of larger surface area of titanium dioxide for photocatalytic reactions.



Fig. 3. The XRD pattern of TiO<sub>2</sub> coated film.



Fig. 4. SEM image of TiO<sub>2</sub> coated film.

#### **3.2. Degradation of MERO**

Total four modes were studied for MERO degradation which includes UV,  $UV/H_2O_2$ ,  $UV/TiO_2$  and  $UV/TiO_2/H_2O_2$  system. Fig. 5 and 6 shows the results for these modes of MERO degradation.

The decrease in MERO concentration was recorded with respect to time as shown in Fig. 5. UV,  $UV/H_2O_2$ ,  $UV/TiO_2$  and  $UV/TiO_2/H_2O_2$  system shows a degradation of 41.14, 85.65, 78.52 and 99.91% respectively during the first hour of irradiation. UV,  $UV/H_2O_2$  and  $UV/TiO_2$  system take about three hours of UV-irradiation to give degradation in the range of 90%. In case of  $UV/H_2O_2$  and  $UV/TiO_2/H_2O_2$  system, a sudden degradation of about 80% was observed within the first half hour of irradiation. It was occurred due to the fast chemical oxidation of MERO with the help of strong oxidizing agent, hydrogen peroxide.







Fig. 6.Percentage degradation as function of time for different modes of degradation.

The degradation efficiency was found to be higher for the UV/TiO<sub>2</sub> and UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> treatment of MERO as compared to other studied treatment. Fig. 5 and 6 clearly shows that the UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> treatment is the most efficient treatment method for antibiotic degradation. Nevertheless, UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system allows the elimination of an equivalent quantity of MERO with in 1hour, where the abatement reaches 99% approximately. It clearly supports the higher economy and effectiveness of this system for antibiotic degradation [54].

Chemical oxidation of MERO antibiotic through hydrogen peroxide alone gives 67.86% degradation, which is not sufficient for resistance prevention in pathogens.

# 3.3 Kinetics of the degradation of MERO

The kinetics of photocatalytic and photolytic degradation of MERO was also studied. The application of the Langmuir-Hinshelwood model for the photocatalytic degradation of MERO was confirmed by the straight line obtained from the representation of the  $\ln(C_0/C)$  versus time plots. Fig. 7 shows the plot of  $ln(C_o/C)$  versus Irradiation time (t) for UV, UV/H<sub>2</sub>O<sub>2</sub>, UV/TiO<sub>2</sub> and UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> processes of MERO degradation. The linearity of the plots as shown in Fig. 7 suggests that the photocatalytic and photolytic degradation follows the pseudo-first order kinetics. Degradation of MERO exhibited the highest rate constant 7.095 h<sup>-1</sup> for UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system followed by 2.245  $h^{-1}$  for UV/TiO<sub>2</sub>, 1.038  $h^{-1}$  for UV/H<sub>2</sub>O<sub>2</sub> and 0.719  $h^{-1}$  for UV system.



Fig. 7.Kinetic analysis for different modes of MERO degradation.

The values of half-life time for the degradation of MERO through UV, UV/H<sub>2</sub>O<sub>2</sub>, UV/TiO<sub>2</sub> and UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system were calculated and tabulated in Table 1. The half-life time values for UV, UV/H<sub>2</sub>O<sub>2</sub>, UV/TiO<sub>2</sub> and UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system was 0.964 h, 0.668 h, 0.309 h and 0.098, respectively. The lowest half-life time and highest reaction constant for the degradation of MERO were obtained for UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system which is an indicative of highest photocatalytic activity.

 Table 1.Half-life time and rate constant for different modes of degradation.

Mode of Degradation	Rate Constant, h <sup>-1</sup>	Half- life time, h
UV	0.719	0.964
UV/H <sub>2</sub> O <sub>2</sub>	1.038	0.668
UV/TiO <sub>2</sub>	2.245	0.309
UV/TiO <sub>2</sub> /H <sub>2</sub> O <sub>2</sub>	7.095	0.098

#### **IV. CONCLUSIONS**

Degradation of MERO has been successfully conducted by photocatalytic process using an immobilized TiO<sub>2</sub> catalyst. It had been additionally compared with photolysis and chemical oxidation using  $H_2O_2$ . It is a promising and Eco-friendly technique; its advantage is that it does not require any kind of separation or filtration prior to treatment, compared to other treatment methods.The degradation potency was found to be maximum for the UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system followed by UV/TiO<sub>2</sub> system among the studied methods.Highest

photocatalytic activity is shown by UV/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> system. The kinetics obeys Langmuir-Hinshelwood model from which kinetic constant were obtained. It shows that the degradation of MERO follows the pseudo-first order kinetics. For identical UV dose, the rate constants were greatly increases with increasing  $H_2O_2$  concentration.

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#### VI. ACRONYMS

- AOPs Advance oxidation processes
- BPR Batch Photocatalytic Reactor
- MERO Meropenem
- SEM Scanning electron microscopy
- UV Ultra-violet light
- XRD X-ray diffraction

#### REFERENCES

- [1] E. Gupta, S. Mohanty, S. Sood, B. Dhawan, B.K. Das and A. Kapil, *Emerging resistance* to carbapenems in a tertiary care hospital in north India, Indian Journal of Medical Research, 124(1), 2006, 95-98.
- [2] N. Gupta, B.M. Limbago, J.B. Patel and A.J. Kallen, Carbapenem-resistant Enterobacteriaceae: epidemiology and prevention, Clinical infectious diseases: an official publication of the Infectious Diseases Society of America 53 (1), 2011, 60–67.
- [3] N. Jaggi, P. Sissodia and L. Sharma, Impact of Antimicrobial Stewardship Programme on Carbapenem Resistance in Gram Negative Isolates in an Indian Tertiary Care Hospital, American Journal of Infectious Diseases, 8(2), 2012, 106-111.
- [4] A. Al-Ahmad , F.D. Daschner and K. Kümmerer, Biodegradability of cefotiam, ciprofloxacin, meropenem, penicillin G and sulfamethoxazole, and inhibition ofwastewater bacteria, Archives of Environmental Contamination and Toxicology, 37, 1999, 158-163.
- [5] K. Kümmerer , A. Al-Ahmad and V. Mersch-Sundermann, Biodegradability of some antibiotics, elimination of their genotoxicity and affection of wastewater bacteria in a simple test, Chemosphere, 40, 2000, 701–710.
- [6] R. Alexy, T. Kümpel and K. Kümmerer, *Assessment of degradation of 18 antibiotics*

*in the Closed Bottle Test, Chemosphere, 57,* 2004, 505–512.

- [7] B. Halling-Sørensen, S. Nors Nielsen, P.F. Lanzky, F. Ingerslev, H.C. Holten Lu<sup>¬</sup> tzhoft and S.E. Jørgensen, Occurrence, fate and effects of pharmaceutical substances in the environment—a review, Chemosphere, 36, 1998, 357–393.
- [8] C. Hartig, T. Storm and M. Jekel, Detection and identification of sulphonamide drugs in municipal waste water by liquid chromatography coupled with electrospray ionisation mass spectrometry, Journal of Chromatography A, 845, 1999, 163–173.
- [9] R. Hirsh, T. Terns, K. Haberer and K.L. Kratz, Occurrence of antibiotics in the aquatic environment, Science of the Total Environment, 225, 1999, 109–118.
- [10] T. Herberer, Occurrence, fate, and removal of pharmaceutical residues in the aquatic environment: a review of recent research data, Toxicology Letters, 131, 2002, 5–17.
- [11] K.H. Choi, Y.H. Kim, J.I. Park, C.K. Park, M.Y. Kim, H.S. Kim, et al., Seasonal variations of several pharmaceutical residues in surface water and sewage treatment plants of Han River, Korea, Science of the Total Environment, 405, 2008, 120–128.
- [12] L. Wollenberger, B. Halling-Sørensen and K.O. Kusk, Acute and chronic toxicity of veterinary antibiotics to Daphnia magna, Chemosphere, 40, 2000, 723–730.
- [13] B. Halling-Sørensen, G. Sengeløv and J. Tjørnelund, *Toxicity of tetracyclines and tetracycline degradation products to environmentally relevant bacteria including selected tetracycline-resistant bacteria, Archives of Environmental Contamination and Toxicology, 42, 2002, 263–271.*
- [14] M.V. Walter and J.W. Vennes, Occurrence of multiple-antibiotic-resistant enteric bacteria in domestic sewage and oxidation lagoons, Applied and Environmental Microbiology, 50(4), 1985, 930–933.
- [15] K. Kümmerer and A. Henninger, Promoting resistance by the emission of antibiotics from hospitals and households into effluents, Clinical Microbiology and Infection, 9, 2003, 1203–1214.
- [16] A. Pruden, R. Pei, H. Storteboom and K.H. Carson, Antibiotic resistance genes as emerging contaminants: studies in Northern Colorado, Environmental Science and Technology, 40, 2006, 7445–7450.
- [17] H. Sørum, Antimicrobial resistance in bacteria of animal origin (F.M. Aarestrup,

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American Society for Microbiology Press, Washington, DC, 2006).

- [18] Q. Zhang, G. Lambert, D. Liao, H. Kim, K. Robin, C. Tung, N. Pourmand and R.H. Austin, Acceleration of Emergence of Bacterial Antibiotic Resistance in Connected Microenvironments, Science 23:333(6050), 2011, 1764-1767.
- [19] J.M. Rolain, R. Canton and G. Cornaglia, Emergence of antibiotic resistance: need for a new paradigm, Clinical Microbiology and Infection, 18(7), 2012, 615–616.
- [20] B. Sun, M. Set and J.S. Clements, Optical study of active species produced by a pulsed corona discharge in water, Journal of Electrostatics, 39, 1997, 189–202.
- [21] A. Lopez, A. Bozzi, G. Mascolo and J. Kiwi, Kinetic investigation on UV and UV/H<sub>2</sub>O<sub>2</sub> degradations of pharmaceutical intermediates in aqueous solution, Journal of Photochemistry and Photobiology A, 156, 2003, 121–6.
- [22] D. Vogna, R. Marotta, A. Napolitano, R. Andreozzi and M. d'Ischia, Advanced oxidation of the pharmaceutical drug diclofenac with UV/H<sub>2</sub>O<sub>2</sub> and ozone, Water Research, 38, 2004, 414–422.
- [23] M. Addamo, V. Augugliaro, A. Di Paola, E. Gracía-López, V. Loddo, G. Marcí and L. Palmisano, *Removal of drugs in aqueous* systems by photoassisted degradation, Journal of Applied Electrochemistry, 35, 2005, 765-774.
- [24] K. Ikehata, N.J. Naghashkar and M.G. El-Din, Degradation of aqueous pharmaceuticals by ozonation and advanced oxidation processes: a review, Ozone: Science & Engineering, 28, 2006, 353-14.
- [25] F. Yuan, C. Hu, X. Hu, J. Qu and M. Yang, Degradation of selected pharmaceuticals in aqueous solution with UV and UV/H<sub>2</sub>O<sub>2</sub>, Water Research, 43, 2009, 1766–1774.
- [26] C. Adams, M. Asce, Y. Wang, K. Loftin and M. Meyer, *Removal of antibiotics from* surface and distilled water in conventional water treatment processes, Journal of Environmental Engineering, 128, 2002, 253-260.
- [27] I. ArslanAlaton and S. Dogruel, Pretreatment of penicillin formulation effluent by advanced oxidation processes, Journal of Hazardous Materials B, 112, 2004, 105-113.
- [28] P. Calza, C. Medana, M. Pazzi, C. Baiocchi and E. Pelizzetti, *Photocatalytic* transformations of sulphonamides on titanium dioxide, Applied Catalysis B, 53, 2004, 63-69.

- [29] S. Kaniou, K. Pitarakis, I. Barlagianni and I. Poulios, *Photocatalytic oxidation of sulfamethazine*, *Chemosphere*, 60, 2005, 372-380.
- [30] W. Baran, J. Sochacka and W. Wardas, *Toxicity and biodegradability of sulfonamides and products of their photocatalytic degradation in aqueous solutions, Chemosphere* 65, 2006, 1295-1299.
- [31] C. Reyes, J. Férnandez, J. Freer, M.A. Mondaca, C. Zaror, S. Malato and H.D. Mansilla, Degradation and inactivation of tetracycline by TiO<sub>2</sub> photocatalyst, Journal of Photochemistry and Photobiology A, 184, 2006, 141-146.
- [32] M.N. Abellán, B. Bayarri, J. Giménez and J. Costa, Photocatalytic degradation of sulfamethoxazole in aqueous suspension of TiO<sub>2</sub>, Applied Catalysis B, 74, 2007, 233-257.
- [33] L. Hu, P.M. Flanders, P.L. Miller and T.J. Strathmann, Oxidation of sulfamethoxazole and related antimicrobial agents by TiO<sub>2</sub>photocatalysis, Water Research, 41, 2007, 2612-2626.
- [34] A. Chatzitakis, C. Berberidou, I. Paspaltsis, G. Kyriakou, T. Slaviadis and I. Poulios, *Photocatalytic degradation and drug activity reduction of Chloramphenicol*, *Water Research*, 42, 2008, 386-394.
- [35] S. Jiao, S. Zheng, D. Yin, L. Wang and L. Chen, Aqueous photolysis of tetracycline and toxicity of photocatalytic products to luminescent bacteria, Chemosphere, 73, 2008, 377-382.
- [36] R. Palominos, J. Freer, M.A. Mondaca and H.D. Mansilla, Evidence for hole photocatalytic participation during oxidation of the antibiotic flumequine, Journal of**Photochemistry** and Photobiology A, 193, 2008, 139-145.
- [37] J. Shaojun, Z. Shourong, Y. Daqiang, W. Lianhong and C. Liangyan, Aqueous oxytetracycline degradation and the toxicity change of degradation compounds in photoirradiation process, Journal of Environmental Sciences, 20, 2008, 806-813.
- [38] I. Kim, N. Yamashita and H. Tanaka, Performance of UV and UV/H<sub>2</sub>O<sub>2</sub> processes for the removal of pharmaceuticals detected in secondary effluent of a sewage treatment plant in Japan, Journal of Hazardous Materials, 166, 2009, 1134-1140.
- [39] R. Palominos, M.A. Mondaca, A. Giraldo, G. Peñeuela, M. Pérez-Moya and H.D. Mansilla, *Photocatalytic oxidation of the antibiotic tetracycline on TiO<sub>2</sub> and*

ZnOsuspensions, Catalysis Today, 144, 2009, 100-105.

- [40] L. Rizzo, S. Meric, M. Guida, D. Kassinos and V. Belgiorno, *Heterogenousphotocatalytic degradation* kinetics and detoxification of an urban wastewater treatment plant effluent contaminated with pharmaceuticals, Water Research, 43, 2009, 4070-4078.
- [41] A.G. Trovó, R.F.P. Nogueira, A. Agüera, C. Sirtori and A.R. Fernández-Alba, Photodegradation of sulfamethoxazole in various aqueous media: persistence, toxicity and photoproducts assessment, Chemosphere, 77, 2009, 1292-1298.
- [42] N.P. Xekoukoulotakis, C. Drosou, C. Brebou, E. Chatzisymeon, E. Hapeshi, D. Fatta-Kassinos and D. Mantzavinos, Kinetics of UV-A/TiO<sub>2</sub> photocatalytic degradation and mineralization of the antibiotic sulfamethoxazole in aqueous matrices, Catalysis Today, 161, 2010, 163-168.
- [43] E. Elmolla and M. Chaudhuri, Degradation of amoxicillin, *ampicillin and cloxacillin antibiotics in aqueous solution by the* UV/ZnOphotocatalytic process, Journal of Hazardous Materials, 173, 2010, 445-449.
- [44] E. Hapeshi, A. Achilleos, M.I. Vasquez, C. Michael, N.P. Xekoukoulotakis, D. Mantzavinos and D. Kassinos, Drugs degrading photocatalytically: kinetics and mechanisms of ofloxacin and atenolol removal on titania suspensions, Water Research, 44, 2010, 1737-1746.
- [45] D. Klauson, J. Babkina, K. Stepanova, M. Krichevskaya and S. Preis, Aqueous photocatalytic oxidation of amoxicillin, Catalysis Today, 151, 2010, 39-45.
- [46] I. Michael, E. Hapeshi, C. Michael and D. Fatta-Kassinos, Solar Fenton and solar TiO<sub>2</sub> catalytic treatment of ofloxacin in secondary treated effluents: evaluation of operational and kinetic parameters, Water Research, 44, 2010, 5450-5462.
- [47] E. Zuccato, S. Castiglioni, R. Bagnati, M. Melis and R. Fanelli, Source, occurrence and fate of antibiotics in the Italian aquatic environment, Journal of Hazardous Materials, 179, 2010, 1042-1048.
- [48] C.C. Ryan, D.T. Tan and W.A. Arnold, Direct and indirect photolysis of sulfamethoxazole and trimethoprim in wastewater treatment plant effluent, Water Research, 45, 2011, 1280-1286.
- [49] F. Yuan, C. Hu, X. Hu, D. Wie, Y. Chen and J. Qu, *Photodegradation and toxicity changes of antibiotics in UV and*

*UV/H*<sub>2</sub>*O*<sub>2</sub>*process, Journal of Hazardous Materials, 185, 2011, 1256-1263.* 

- [50] E. Elmolla and M. Chaudhuri, *The* feasibility of using combined *TiO*<sub>2</sub>photocatalysis-SBR process for antibiotic wastewater treatment, Desalination, 272, 2011, 218-224.
- [51] N. Negishi, K. Takeuchi and T. Ibusuki, Preparation of the TiO<sub>2</sub> thin film photocatalyst by Dip-Coating process, The Journal of Sol-Gel Science and Technology, 13, 1998, 691-694.
- [52] A.W. Bauer, W.M.M. Kirby, J.C. Sherrisn and M. Turk, Antibiotic susceptibility testing by a standardized single disc method, American Journal of Clinical Pathology, 45, 1966, 493-496.
- [53] T. Ohno, K. Sarukawa, K. Tokieda and M. Matsumura, Morphology of a TiO<sub>2</sub> Photocatalyst (Degussa, P25) Consisting of Anatase and Rutile Crystalline Phases, Journal of Catalysis, 203, 2001, 82-86.
- [54] E. Elmolla and M. Chaudhuri, Photocatalytic degradation of amoxicillin, *ampicillin and cloxacillin antibiotics in aqueous solution using UV/TiO<sub>2</sub> and UV/H<sub>2</sub>O<sub>2</sub>/TiO<sub>2</sub> photocatalysis, Desalination*, 252, 2010, 46-52.